An asthmatic child with allergic bronchopulmonary aspergillosis (ABPA)

Öner Özdemir
Division of Allergy and Immunology, Department of Pediatrics, Research and Training Hospital of Sakarya University, Adapazarı, Sakarya, Turkey. E-mail: ozdemir_oner@hotmail.com
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Allergic bronchopulmonary aspergillosis (ABPA) results from a TH2 hypersensitivity lung disease in response to bronchial colonization of Aspergillus fumigatus in the respiratory tract. ABPA is a very rare disease in the pediatric population; the prevalence is anticipated to be about 1–2% in adult asthma patients and 2–15% in cystic fibrosis patients. In pediatric asthma patients, only rare case reports on ABPA have been published. We present a case of a 12-year-old boy admitted to our hospital with worsening (mild persistent) asthma symptoms such as a recurrent cough, episodic wheezing, and dyspnea for the last two years. He was diagnosed with pneumonia several times and suspected of having tuberculosis during this time. Chest X-ray showed bilaterally perihilar opacities and computed tomography findings revealed central bronchiectasis. With the clinical and laboratory findings, the patient fulfilled 7/8 major diagnostic criteria for ABPA. Treatment with oral corticosteroids and itraconazole was effective. To the best of our knowledge, this is the first reported case of pediatric ABPA with previously documented asthma from Turkey.

Key words: allergic bronchopulmonary aspergillosis, asthma, central bronchiectasis, aspergillus fumigatus.

Allergic bronchopulmonary aspergillosis (ABPA) is a T-helper 2 (TH2) hypersensitivity reaction to fungus, known as Aspergillus fumigatus (AF), allergens colonizing the bronchial tree.1,2 Although ABPA is infrequently seen in children, it is more common in children with cystic fibrosis (CF).3,4 ABPA is predominantly seen in adult asthmatics, especially corticosteroid dependent asthmatics, in the third and fourth decades of life. Its prevalence varies from 1% to 25% of persistent asthma patients in Europe.5-7 In 1985, pediatric persistent asthma patients were evaluated for ABPA by Chetty et al.8 and the prevalence was demonstrated to be 15%. There are also probably significant regional differences in ABPA prevalence, with a lower rate likely in the USA and higher rate likely in India.7,9 Furthermore, Denning et al.10 have estimated that the global prevalence of ABPA may be 0.7-3.5% of patients with asthma.

The pathogenesis of ABPA is not fully known. Several type of immune responses seem to be implicated, including AF-specific IgE-mediated type I, specific IgG-mediated type III hypersensitivity reactions, and abnormal T-lymphocyte responses.11 There are some genetic mutations like single-nucleotide polymorphisms and expression of some human leukocyte antigen (HLA) blamed for ABPA susceptibility. Chauhan et al.12 reported that asthmatic patients expressing HLA-DR2 and/or DR5 and possibly HLA-DR4 or DR7 and lacking HLA-DQ2 were at amplified risk to have ABPA after exposure to AF.3

ABPA diagnosis might be easily overlooked or delayed, because its clinical picture is often impossible to differentiate from those of...
more frequent lung disorders e.g. uncontrolled asthma, pneumonia and tuberculosis. ABPA is distinguished by exacerbations of asthma, worsening of pulmonary function, recurrent transient or fixed pulmonary infiltrates, peripheral blood and pulmonary eosinophilia and elevated total IgE level. Here, we report a 12-year-old boy who was investigated for recurrent cough, pneumonia, episodic wheezing (worsening mild persistent asthma symptoms) and later diagnosed as ABPA with underlying asthma.

**Case Report**

A 12-year-old boy [height: 1.38 cm (3-10p), weight: 28 kg (<3p)] was referred to our department due to worsening asthma symptoms (e.g. intractable cough, episodic wheezing and dyspnea) for two years and suspicion of tuberculosis. In his past medical history, he was previously treated for bronchopneumonia a couple of times and montelukast, inhaled corticosteroids (fluticasone inhaler, 125 μg, bid) and salbutamol were prescribed for mild persistent asthma since he had been diagnosed with asthma at 9 years of age. Although his cough and symptoms regressed somewhat with the treatment, he did not improve entirely. While he was investigated for uncontrolled asthma symptoms, bilateral perihilar opacities (Fig. 1a) on chest X ray (CXR) and central bronchiectasis on his computed tomography (CT) (Fig. 2a-c) were detected. In his family history, nothing was significant.

During the course of admission, respiratory rate was 20 per minute; oxygen saturation measured by pulse oxymetry was 96% and body temperature was 37°C. The patient appeared to be healthy with no signs of respiratory distress. Auscultation of the chest revealed no ronchi or crackles. Further physical examination demonstrated no other pathological findings. Complete blood cell count showed white blood cells: 8.500/mm³ (neutrophils, 30.6%; lymphocytes, 42.2%;

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**Rosenberg-Patterson Criteria**

<table>
<thead>
<tr>
<th>Our case (the patient)</th>
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<tbody>
<tr>
<td>1) FEV1:63%; FVC:52%</td>
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<tr>
<td>2) FEV1:83%; FVC:71%</td>
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<tr>
<td>(reversibility+)</td>
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<td>135 mg/L / 38.6 KU/L</td>
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**Table I.** Comparison of Laboratory and Clinical Findings of Our Patient with Rosenberg-Patterson Criteria.

<table>
<thead>
<tr>
<th>Rosenbberg-Patterson Criteria</th>
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<tr>
<td>Major criteria 1,2</td>
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<tr>
<td>1. Asthma (spirometric) deterioration not attributable to another etiology</td>
<td>1-) FEV1:63%; FVC:52%</td>
</tr>
<tr>
<td>2. Skin prick test to <em>Aspergillus fumigatus</em></td>
<td>Positive</td>
</tr>
<tr>
<td>3. Elevated serum specific IgG and IgE to <em>Aspergillus fumigatus</em></td>
<td>135 mg/L / 38.6 KU/L</td>
</tr>
<tr>
<td>Specific IgE greater than twice non-ABPA IgE A. fumigatus-positive asthmatics</td>
<td></td>
</tr>
<tr>
<td>4. Serum precipitating IgG antibodies (precipitins) to <em>Aspergillus fumigatus</em></td>
<td>N/A</td>
</tr>
<tr>
<td>5. Elevated total serum IgE level ≥1000 (or ≥417) IU/ml</td>
<td>1.986 IU/ml</td>
</tr>
<tr>
<td>6. Peripheral blood eosinophilia (≥1.000 cells/μL) (fleeting shadows)</td>
<td>%20.2 (1.720/mm³)</td>
</tr>
<tr>
<td>7. Chest radiographic transient or fixed lung infiltrate(s) (fleeting shadows)</td>
<td>Positive</td>
</tr>
<tr>
<td>8. Lung CT: Central (proximal) bronchiectasis in the upper lobes</td>
<td>Positive</td>
</tr>
<tr>
<td>Minor criteria</td>
<td></td>
</tr>
<tr>
<td>10. Delayed type (type III, Arthus type) skin reaction to Asp. antigens</td>
<td>N/A</td>
</tr>
<tr>
<td>11. Presence of Aspergillus in sputum</td>
<td>Negative</td>
</tr>
<tr>
<td>12. Golden (brownish black) sputum /mucus plaques</td>
<td>Positive</td>
</tr>
</tbody>
</table>

(1) Criteria 1–8, ABPA-central bronchiectasis, ABPA-CB; (2) Criteria 1–7, ABPA-seropositive, ABPA-S. ABPA: Allergic bronchopulmonary aspergillosis, N/A= not available, FEV1= forced expiratory volume in 1 second, FVC= forced vital capacity.
monocytes, 6.18%; eosinophils, 20.2% (1.720/mm$^3$); basophils, 0.80%); hemoglobin, 12.5 g/dl; and platelet, 274,000/mm$^3$. ESR was 9 mm/hr. The C-reactive protein was <3.14 mg/L. Routine biochemistry was within normal range. Tuberculosis skin (PPD) test was negative. Serum total IgE level was 1.986 IU/ml. Pulmonary function tests in our outpatient clinic revealed moderate obstructive pattern with positive reversibility (forced vital capacity [FVC]: 52%, 71%; forced expiratory volume in 1 second [FEV1]: 63%, 83%; FEV1/FVC: 115%, 114% (pre- and post-bronchodilation values; respectively). According to modified Rosenberg-Patterson criteria, diagnosis of ABPA was made with positive 7 out of 8 major criteria (Table I). Those 7 major criteria met by the patient were perihilar opacities, central bronchiectasis, eosinophilia, high total IgE, specific IgE (38.6 KU/L) and IgG antibodies (135mg/L) to AF, twice positive skin prick test for AF (wheal of 4×4 mm) and reduced FEV1/FVC on spirometry. Galactomannan antigen was found to be negative. Sputum culture, PCR, T-spot® and Quantiferon® tests for Mycoplasma tuberculosis were negative. Sweat test for CF was negative twice in our and another institution. The entire coding regions of 27 exons of the cystic fibrosis transmembrane conductance regulator gene (CFTR) were amplified by polymerase chain reaction (PCR) using specific primers and the DNA sequencing results were normal. Bronchoscopy showed mucus plugging in the airways and bronchoalveolar lavage (BAL) fluid examined. Cytology of the BAL mostly showed leukocytosis (62% neutrophilic granulocytes) and gram negative bacillus. Pseudomonas aeruginosa was isolated from the BAL fluid culture and ceftazidime treatment (150mg/kg/day) was given for 14 days. ANA and ANCA test results were negative. Serum humoral antibody levels (IgG, IgA, IgM and IgG1- IgG2- IgG3- IgG4) were normal. T-cell subtypes were also detected to be within normal limits.

Intravenous prednisolone (1 mg/kg/day, 30 mg/day for 2 weeks) and oral itraconazole (200 mg/day, 4 months) treatments were started. After 14 days of oral prednisolone (1 mg/kg/day) continued on alternate day for 2 months, it was tapered 4 mg down every other week for 3 months and stopped at 4th months. After
4 months, oral itraconazole reduced to 100 mg/day and continued for another 4 months and discontinued at the end of 8th month. During steroid therapy his plasma glucose levels increased and remained high at the 6th week of treatment. All hormonal evaluations including insulin, C-peptid, cortizole, prolan, tiroid hormones, amylase and lipase were found to be normal. Diagnosis of steroid-induced diabetes mellitus was made by our endocrinologist and insulin treatment had to start. After tapering steroid dose down, his blood glucose levels decreased and remained within normal ranges. In addition to high glucose levels, mild-moderate hirsutism developed. After 14 days of admission, he was discharged. During follow-up his asthma has remained under control with the maintenance therapy of montelukast (5 mg, qd) and budesonid 80 μg+formoterol fumarat dihidrat 4.5 μg (bid). Under this treatment, his symptoms resolved day by day. At follow-up, control chest radiograph (Fig. 1b) showed improvement on bilateral perihilar opacities and central bronchiecatic lesions seemed to resolve on control CT (Fig. 2d,e). During and after the treatment, patient’s total serum IgE decreased gradually (Fig. 3). Informed consent was obtained from patient’s legal guardian (parents).

Discussion

The clinical presentation of ABPA varies from mild bronchospasm (asthma) to fibrotic parenchymal lung disease. Although very few authors have reported ABPA development in pediatric asthma patients, it could be suspected in uncontrolled asthma that might be related with fungal sensitzation. If undetected or inadequately treated, ABPA causes destruction in respiratory tract, bronchiectasis, and/or lung fibrosis, consequently considerable morbidity and mortality. Our patient’s worsening clinical symptoms and pulmonary functions suggesting uncontrolled asthma was detected to be associated with bronchiectasis induced by ABPA.

The definite diagnosis of ABPA is not simple due to overlapping clinical and radiological signs. ABPA diagnosis is made by a group of clinical, laboratory, and radiographic findings including asthma, elevated total IgE level, blood eosinophilia, evidence of sensitzation to AF, recurrent pulmonary parenchymal infiltrates and bronchiectasis. The classic Rosenberg–Patterson criteria are most often utilized for the ABPA diagnosis in non-CF patients. The existence of six out of the eight major criteria makes the diagnosis almost certain. In a 2012 review, the minimal criteria necessary for the diagnosis of ABPA were defined as: (1) asthma or CF with worsening of lung function, (2) skin prick test positivity to Aspergillus species, (3) total serum IgE level of 1.000 ng/ml (416 IU/ml) or higher, (4) amplified Aspergillus species-specific IgE and IgG antibodies, and (5) chest radiographic infiltrates. Additional criteria consist of peripheral blood eosinophilia, Aspergillus species serum precipitating antibodies, central bronchiectasis, and Aspergillus species–containing mucus plugs. Our patient fulfilled 7/8 classic major criteria for ABPA diagnosis (Table 1). In the absence of typical proximal (central) bronchiectasis, the disorder is called seropositive ABPA (ABPA-S) in place of ABPA with central bronchiectasis (ABPA-CB). Our patient had ABPA-CB.

Although ABPA is known to occur mostly with underlying asthma and CF, there have been several published reports on ABPA development without previously diagnosed underlying disease such as asthma. Although our patient was also treated for pneumonia several times after being diagnosed and treated for asthma, on the other hand, clinical and CXR findings suggested tuberculosis infection in our patient. Tuberculosis is somewhat of a concern in Turkey; therefore the patient was admitted for tuberculosis work-up and further investigation. Tuberculosis should be considered in the differential diagnosis of ABPA. This is apparent in a study, as 38.4% of ABPA patients had a record of anti-tuberculosis treatment. However, tuberculosis was ruled out by radiologic and laboratory evaluations in our patient. With the clinical and laboratory findings, our patient was considered to have uncontrolled asthma. Key point in the diagnosis of ABPA is suspicion and ABPA should be considered in patients with uncontrolled asthma symptoms. During further investigation, his spirometry showed moderate obstruction with positive reversibility and then the diagnosis of asthma was confirmed. Like our case, diagnosis of ABPA might be delayed because of clinical signs and symptoms mimicking recurrent pneumonia or tuberculosis. In our
search of the literature from Turkey, we have come across just one case report of 3 adult asthmatic ABPA cases by Erkekol et al.\textsuperscript{21} To the best of our knowledge, this will be the first reported case of ABPA with underlying pediatric asthma from Turkey.

CFTR gene mutations have been reported to be increased in asthmatic patients who developed ABPA. In a meta-analysis, Agarwal et al.\textsuperscript{22} also demonstrated a higher incidence of heterozygous CFTR mutations in asthmatic ABPA patients. However, in our patient DNA sequence analysis for CFTR mutations was normal. In the literature, early colonization of the respiratory tract with \textit{Pseudomonas aeruginosa} was considered to be related with the development of ABPA in younger patients.\textsuperscript{4} In our patient \textit{Pseudomonas aeruginosa} was isolated from the BAL fluid culture and we were not sure whether this colonization was associated with the development of ABPA or not. Patterson et al.\textsuperscript{23} categorized ABPA into 5 stages dependent on asthma severity and radiographic or laboratory findings. In this classification, stage I is the early acute phase of ABPA. In stage II, the disease in remission; the infiltrates clear, symptoms decrease and the serum IgE value reduces but usually remains elevated. It looks like our patient is currently at stage II.

Some authors presume that the ABPA is missed in numerous cases with reports of mean diagnostic latency of even 10 years between the beginning of symptoms and the diagnosis.\textsuperscript{24} In our case, the delay between the onset of symptoms and the diagnosis was just a couple of years. There are 2 essential causes for the delay in ABPA diagnosis: the disorder seems to be uncommon and unspecific symptoms can be simply misunderstood as exacerbations of an underlying disease.

ABPA should be suspected in an asthmatic patient if they have the following: (1) new or recent infiltrate(s) implying pneumonia or tuberculosis on lung radiography or thorax CT that have not cured with antibiotics and standard physiotherapy, (2) clinical and laboratory findings suggesting uncontrolled asthma despite adequate therapy, (3) unresolved pneumonia and bronchiectasis irrespective of history of asthma.\textsuperscript{19} Moreover, in the differential diagnosis of ABPA, pulmonary exacerbations by bacterial or viral agents; CF, asthma; atopy; severe small airways disease; gastroesophageal reflux aspiration; pneumothorax; hyperimmunoglobulin E and Churg-Strauss syndrome should be ruled out. Since our patient fulfilled 7/8 diagnostic major criteria suggested by Rosenberg–Patterson,\textsuperscript{14} the other conditions were easily ruled out.

Management of ABPA is based on clinical symptoms. Main goals of treatment are reduction of pulmonary inflammation, prevention of progression of the disease and controlling exacerbations.\textsuperscript{16} Treatment essentially consists of systemic corticosteroid with/without itraconazole.\textsuperscript{25} Oral corticosteroids are the therapy of choice for ABPA. Oral itraconazole treatment is still controversial. Antifungal agents like itraconazole are generally used in CF-ABPA patients; it is thought to diminish the fungal load of the respiratory tract.\textsuperscript{26} There is some evidence that itraconazole has a steroid sparing effect and potentially prevent the recurrence of ABPA.\textsuperscript{27} A recently reported clinical study showed that even refractory pediatric asthma and children sensitized to fungal antigen might benefit from adjunct treatment with itraconazole. Some of the published reports also have shown beneficial effects of combination treatment of itraconazole with prednisolone daily compared with only prednisolone in terms of controlling exacerbations, reduced sputum production, reduced total IgE, better remission of radiological findings and improved FEV1.\textsuperscript{28}

Follow-up of the patient could be done by observing clinical symptoms and total IgE levels. Reducing total IgE levels 35–50\% of the value at the time of diagnosis is expected after starting treatment.\textsuperscript{26} In addition, these patients should be evaluated for adverse effects of long term systemic steroid and itraconazole use. Our case has been successfully treated with combination treatment of methylprednisolone and itraconazole. Clinical symptoms and radiological findings improved, and also total IgE levels decreased. Although these were satisfactory results, he experienced uncontrolled high blood glucose levels caused by steroid-induced diabetes mellitus as a side effect. Fortunately, besides using insulin, tapering the steroid dose down and eventually stopping the steroid brought his blood glucose down to a normal level.
In conclusion; although it is a rare disease, the physicians should keep ABPA in mind in pediatric patients presenting with uncontrolled asthma despite adequate therapy, unresolved recurrent pneumonia and in those complaining of intractable cough. Also evaluating serum total IgE levels and blood eosinophil levels are high. Early and correct ABPA diagnosis and aggressive treatment can diminish needless antibiotic treatments and prevent development of irreversible fibrotic lung damage.

REFERENCES
